PART III

CLINICAL STUDIES
Activated protein C attenuates pulmonary coagulopathy in patients with acute respiratory distress syndrome

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**ABSTRACT**

*INTRODUCTION:* Acute respiratory distress syndrome (ARDS) frequently complicates critical illness. We hypothesized that infusion of recombinant human activated protein C (rh-APC), a natural anticoagulant, would attenuate pulmonary coagulopathy and injury.

*METHODS:* In this sub study of a multicenter open-label randomized controlled trial of patients with ARDS, we compared intravenous infusion of rh–APC (24 mcg/kg/hr for 96 hours) with placebo. Patients with sepsis or septic shock were excluded.

*RESULTS:* In 27 patients serial non-directed bronchoalveolar lavage fluid (NBLF) samples were obtained: 16 patients were treated with rh–APC, 11 patients with placebo. Rh–APC infusion was associated with higher APC levels in plasma during the infusion period of 4 days (P = 0.001), as well as higher APC levels in NBLF up to day 5 after start of infusion (P = 0.028). Infusion of rh–APC was associated with lower levels of thrombin–antithrombin complexes (P = 0.009) and soluble tissue factor (P = 0.011) in NBLF, compared to treatment with placebo. Infusion of rh–APC affected fibrinolysis, as plasminogen activator activity levels in NBLF were higher in the patients treated with rh–APC (P = 0.01), presumably due to lower NBLF levels of plasminogen activator inhibitor 1, (P = 0.01). Rh-APC infusion decreased the lung injury score (P = 0.005) and simplified acute physiology score (P = 0.013) on day 5, when compared to baseline. Rh–APC infusion was not associated with bleeding complications.

*CONCLUSION:* Infusion of rh–APC in patients with ARDS attenuates pulmonary coagulopathy and injury.
INTRODUCTION

Acute respiratory distress syndrome (ARDS), and its milder form, previously known as acute lung injury (ALI) [1], occur at rates between 30 and 80 per 100,000 person-years and are a common cause of respiratory insufficiency and admission to the intensive care unit (ICU) [2,3]. ARDS is associated with high morbidity and mortality [2]. Despite efforts to find a specific treatment for ALI/ARDS [4], treatment nowadays is merely supportive and directed at avoiding additional harm (by mechanical ventilation).

Pulmonary coagulopathy, which results from activation of coagulation, defective anticoagulant pathways and inhibition of fibrinolysis, is invariably present in ARDS and is likely to contribute to its pathogenesis [5-8]. In patients with ARDS, pulmonary activated protein C (APC) levels are depressed [8-11]. Infusion of recombinant human (rh) APC was found to reduce mortality of patients with severe sepsis and a high disease severity [12], although this was offset by other trial evidence and a recently published study of patients with septic shock, which even led to market withdrawal of commercial rh–APC (Xigris®) [13]. Of note, in the phase III trial that initially resulted in licensing of Xigris®, administration of rh–APC was particularly effective in patients who presented with severe community–acquired pneumonia as the source of sepsis [14]. Therefore, the beneficial effect of infusion of rh–APC in patients with severe sepsis as found in some studies could, at least in part, be attributed to effects of rh–APC on coagulopathy in the lung [14]. This is supported by results of pre–clinical studies in rats evaluating the effect of infusion of rh–APC in models of ARDS showing anticoagulant effects in the lungs [15,16]. Also, one study in healthy volunteers demonstrated infusion of rh–APC to affect pulmonary coagulation after intrapulmonary challenge with endotoxin [9].

To date, no data exist on the effects of infusion of rh–APC on the pulmonary coagulopathy present in patients with ARDS. We hypothesized that infusion of rh–APC would attenuate this pulmonary coagulopathy, which could be beneficial to patients with ARDS. Therefore, we performed an open–label randomized controlled trial of patients with ARDS comparing intravenous infusion of rh–APC with placebo, with respect to the pulmonary and systemic pro- and anticoagulant balance and organ functions.

MATERIALS AND METHODS

STUDY DESIGN

This is a sub study of INFectious and INFlammatory Acute Lung Injury / Acute Respiratory Distress Syndrome (INFALI), a multicenter open–label randomized controlled trial of patients with ALI/ARDS (trial registration number ISRCTN 52566874). The Ethics Committee of VU University Medical Center, Amsterdam, the Netherlands, approved the study protocol. Written informed consent was obtained from all patients or their next of kin before inclusion into the trial.
**Inclusion and exclusion criteria**

Patients, over 18 years of age and admitted to the mixed medical–surgical ICUs of one of two participating university hospitals, were to be enrolled within 24 hours after ARDS was diagnosed irrespective of the need for (invasive) ventilatory support. ARDS was diagnosed using the North American European Consensus Conference (NAECC) definition [17]. Patients were excluded if rh–APC treatment was indicated based on current national guidelines at the time of the study (i.e., severe sepsis or septic shock). Additional exclusion criteria were: platelet count < 30 x 10⁹/L, any major surgery within 12 hours before inclusion, acute bleeding, severe head trauma, intracranial surgery or stroke within 3 months before inclusion, known intracranial abnormalities (e.g., malignancies or other tumors, arteriovenous malformation), known hypercoagulability (e.g., protein C resistance, hereditary deficiency of protein C, protein S or antithrombin, or anticardiolipin– or antiphospholipid–antibodies), congenital hemorrhagic diathesis, pregnancy or breast feeding, liver cirrhosis with portal hypertension and/or esophageal varices, presence of an epidural catheter; severely immune–compromised status (e.g., HIV–infected patients with CD4 count < 50/mL, and patients treated with immunosuppressive medication following bone marrow or solid organ transplantation). The following concomitant medications were reasons for exclusion: heparin in therapeutic dose (within 8 hours of study entry), coumarin derivatives at any dose (within 7 days of study entry), acetylsalicylic acid at a dose > 650 mg/day (within previous 3 days of study entry), thrombolytic therapy at any dose (within previous 3 days of study entry), glycoprotein IIb/IIIa inhibitors at any dose (within 7 days of study entry), antithrombin at any dose (within 3 days of study entry) and previous treatment with rh–APC (at any time within study entry). Prophylactic dose of low molecular weight heparin was acceptable.

**Treatment protocol**

All patients were treated according to international guidelines, by the discretion of the supervising intensivists. If needed, mechanical ventilation was performed after endotracheal intubation, in a pressure–controlled mode, aiming at a maximum airway pressures < 35 cmH₂O, and tidal volumes ≤ 6 mL/kg predicted ideal body weight, with or without proning. Antibiotic therapy was guided by Gram–stains and cultures, according to local guidelines for antimicrobial therapy. Fluid therapy consisted of crystalloids, with or without gelatins and/or hydroxyethyl starches, in order to maintain arterial blood pressure (MAP > 70 mmHg) and diuresis (> 30 mL/h).

**Treatment assignment**

Patients were randomly assigned to infusion of rh–APC or placebo (normal saline). Prior to the start of the trial sealed opaque envelopes, containing the treatment assignment for each patient, were numbered through block randomization, with 6 blocks of patients.
Activated protein C attenuates pulmonary coagulopathy in patients with acute respiratory distress syndrome

**STUDY PROTOCOL**

Rh–APC (Eli Lilly, Indianapolis, IN, USA), at a dose of 24 mcg/kg/hr, or placebo was infused at a constant rate for a total of 96 hours [12]. Infusion of rh–APC was interrupted 1 hour before any invasive percutaneous procedure or major surgery. When no bleeding complications occurred, infusion of rh–APC was resumed 1 hour after a percutaneous procedure, and 12 hours after major surgery, in line with international guidelines.

**DATA COLLECTION**

Upon enrollment data on baseline demographics, co–morbidity and reasons of admission to the intensive care unit (ICU), as well as hemodynamic and respiratory parameters were collected. The acute physiology, age and chronic health evaluation II score (APACHE II) [18], the simplified acute physiology score (SAPS II) [19], the sequential organ failure assessment score (SOFA) [20] and the lung injury score (LIS) [21] were calculated daily until day 5 and every second day thereafter. The number of ventilator-free days was assessed, defined as the number of days from randomization to day 28 after achieving unassisted breathing for at least 2 consecutive calendar days. If a patient who achieved unassisted breathing, subsequently required additional assisted breathing, and once again achieved unassisted breathing, only the ventilator-free days after beginning the final period of unassisted breathing were counted. Patients who died before day 28, were assigned zero ventilator-free days. Both 28-day, ICU and in-hospital mortality were assessed. Follow up was discontinued after 1 year.

**NON–DIRECTED BRONCHOALVEOLAR LAVAGE AND BLOOD SAMPLING**

Non–directed bronchoalveolar lavage (NBL) was performed by an experienced intensivist by instilling 20 mL of sterile 0.9% saline via a 50 centimeter 14–Gauge tracheal suction catheter as described previously [22]. In short, the distal end of the suction catheter was inserted through the endotracheal tube until resistance was encountered. The 20 mL of sterile 0.9% saline was instilled over 10 to 15 seconds, and (in part) aspirated before withdrawal of the catheter. Volume of aspirated NBL–fluid (F) was at least 10 mL. Before NBL was performed, blood was collected, preferably from an indwelling arterial catheter or central venous catheter. Sometimes peripheral blood sampling was necessary.

**ASSAYS**

NBLF and blood samples were centrifuged at 1500 x g for 15 minutes and the supernatant was stored at –80°C until assays were performed. Levels of APC were determined using an enzyme capture assay using monoclonal antibody HAPC 1555 and chromogenic substrate Spectrozyme Pca (American Diagnostics, Greenwich, CT). Thrombin–antithrombin complexes (TATc) and soluble tissue factor (sTF) were measured using ELISA (TATc; Behring, Marburg, Germany; sTF: American Diagnostics, Greenwich, CT). Antithrombin (AT), plasminogen activator activity (PAA) and plasminogen activator inhibitor (PAI)–1 were measured by automated amidolytic assays [23].
**Statistical Analysis**

Data were checked for distribution with help of the Kolmogorov-Smirnov test. Data were expressed as medians (interquartile ranges), or absolute numbers where appropriate. Nonparametric data were analyzed using Mann–Whitney U, Wilcoxon signed–ranks and Fisher’s exact test. A P–value of < 0.05 was considered statistically significant. Exact P-values are given unless < 0.001. Statistical analysis was performed using SPSS 19.0 (SPSS, Chicago, IL, USA) and Prism 5.0 (GraphPad Software, San Diego, CA, USA).

**Results**

Between 1 January 2007 and 1 May 2011 9,484 patients were assessed for eligibility. Of these patients 1274 were diagnosed with ARDS, of which 1203 were excluded or refused participation in the trial, leaving 71 patients for study inclusion. The most common reasons for the large number of patients excluded from participation was that many patients with ARDS had concurrent organ failure, were concomitantly treated with anticoagulant medication, or had an increased risk of bleeding. A consort diagram is shown in Figure 1. In 44 patients it was not considered safe (prone positioning, high levels of PEEP) or appropriate (not fully sedated) to perform NBL (by the treating intensivist). Patient characteristics, hemodynamic and respiratory baseline values, as well as disease severity scores are shown in Table 1. Groups were comparable with respect to disease severity and pulmonary condition. The most frequent cause of ARDS was pneumonia. In both groups, no bleeding complications or other serious adverse events were observed.

**Fig.1 CONSORT diagram.**

ARDS, acute respiratory distress syndrome; rh-APC, recombinant human activated protein C; SCT, stem cell transplantation; plt, platelets; NBL, non-directed bronchoalveolar lavage
### Table 1. Baseline characteristics

<table>
<thead>
<tr>
<th></th>
<th>Rh-APC (n=16)</th>
<th>Placebo (n=11)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>General</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Age</td>
<td>64 (23)</td>
<td>61.5 (28.5)</td>
<td>.47</td>
</tr>
<tr>
<td>Sex (male)</td>
<td>7</td>
<td>7</td>
<td>.44</td>
</tr>
<tr>
<td>Reason of admission</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>pneumonia</td>
<td>7</td>
<td>8</td>
<td></td>
</tr>
<tr>
<td>abdominal sepsis</td>
<td>4</td>
<td>0</td>
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<tr>
<td>sepsis</td>
<td>5</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>near-drowning</td>
<td>0</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>APACHE 2</td>
<td>14 (8.25)</td>
<td>15 (4)</td>
<td>.46</td>
</tr>
<tr>
<td>SAPS 2</td>
<td>37 (19.5)</td>
<td>34.5 (5.25)</td>
<td>.31</td>
</tr>
<tr>
<td>SOFA</td>
<td>6 (2.5)</td>
<td>5.5 (6)</td>
<td>.44</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td>36.8 (2.25)</td>
<td>36.9 (2.35)</td>
<td>.90</td>
</tr>
<tr>
<td>Heart rate (min)</td>
<td>113 (55)</td>
<td>111 (33.25)</td>
<td>.86</td>
</tr>
<tr>
<td>MAP (mmHg)</td>
<td>75.5 (10.75)</td>
<td>73.5 (22.75)</td>
<td>.79</td>
</tr>
<tr>
<td>Vasopressor</td>
<td>11</td>
<td>7</td>
<td>.16</td>
</tr>
<tr>
<td>Creatinine (mmol/L)</td>
<td>79 (78.75)</td>
<td>86 (111)</td>
<td>.82</td>
</tr>
<tr>
<td><strong>Respiratory</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Respiratory rate (min)</td>
<td>25 (9.75)</td>
<td>21.5 (13.25)</td>
<td>.41</td>
</tr>
<tr>
<td>PaO\textsubscript{2}/F\textsubscript{I}O\textsubscript{2}-ratio</td>
<td>150 (70)</td>
<td>190 (88.25)</td>
<td>.15</td>
</tr>
<tr>
<td>PEEP (cm H\textsubscript{2}O)</td>
<td>11 (4.75)</td>
<td>9.5 (2.75)</td>
<td>.12</td>
</tr>
<tr>
<td>LIS</td>
<td>2.5 (0.75)</td>
<td>2.1 (1)</td>
<td>.16</td>
</tr>
</tbody>
</table>

Data are expressed as medians (interquartile ranges), or absolute numbers where appropriate.

Rh-APC, Recombinant human activated protein C; MAP, mean arterial pressure; PaO\textsubscript{2}, partial arterial oxygen tension; F\textsubscript{I}O\textsubscript{2}, fraction of inspired oxygen; PEEP, positive end expiratory pressure; APACHE, Acute Physiology and Chronic Health Evaluation; SAPS, Simplified Acute Physiology Score; SOFA, Sequential Organ Failure Assessment; LIS, Lung Injury Score

**Systemic anticoagulant effects of infusion of rh–APC**

Infusion of rh–APC was associated with higher APC levels in plasma during treatment (P = 0.001). After discontinuation of infusion of rh–APC, APC levels in plasma became comparable to those in patients receiving placebo (Figure 2). Infusion of rh–APC was associated with lower TATc levels in plasma during and after treatment with rh–APC, and levels remained lower until day 7 (P = 0.014) (Figure 2). Infusion of rh–APC was associated with higher PAA levels in plasma during and after treatment (until day 7) with rh–APC (P = 0.014) (Figure 2), and lower PAI–1 levels in plasma (P = 0.014) (Figure 2).
Fig. 2 Effect of recombinant human activated protein (rh-APC) administration on systemic levels of APC, thrombin-antithrombin complexes (TATc), plasminogen activator activity (PAA) and plasminogen activator inhibitor-1 (PAI-1). Differences in APC were significant on day 2 – 5 (P < 0.01). Differences in TATc, PAA and PAI-1 were significant on day 2 – 5 (P < 0.01) and day 7 (P < 0.05).

PULMONARY ANTICOAGULANT EFFECTS OF INFUSION OF RH–APC

Infusion of rh–APC was associated with increased APC levels in NBLF (P = 0.028), which remained higher than in patients treated with placebo up to the end of the observation period (Figure 3). Infusion of rh–APC was associated with a reduction in the thrombin generation marker TATc in NBLF, during and after infusion of rh–APC (P = 0.009) (Figure 3). However, AT levels in NBLF were similar in both study groups (Figure 3). PAA levels in NBLF were higher in patients treated with rh–APC (P = 0.01) (Figure 3). Similar as in plasma, levels of PAI–1 were lower in NBLF (P = 0.01) (Figure 3). Levels of sTF in NBLF showed a marked decline in patients treated with rh–APC (P = 0.011) (Figure 3).
Effect of recombinant human activated protein (rh-APC) administration on levels of APC, thrombin-antithrombin complexes (TATc), antithrombin (AT), plasminogen activator activity (PAA), plasminogen activator inhibitor-1 (PAI-1) and soluble tissue factor (sTF) in non-directed bronchoalveolar lavage fluid. Differences in APC, PAA, PAI-1 and sTF were significant on day 2 – 4 (P < 0.01) and day 5 (P < 0.05). Differences in TATc were significant on day 2 – 5 (P < 0.01). There were no significant differences in AT.

**CLINICAL PARAMETERS**

In patients treated with rh-APC the baseline LIS (P = 0.005) and SAPS (0.013) had decreased on day 5, with reduced SOFA score following on day 15 (P = 0.046). In patients treated with placebo, there was a decrease in the SOFA score on day 5 (P = 0.047), which did not endure until day 15. The number of ventilator-free days appeared to be less in patients treated with Rh-APC, but statistical significance was not reached. Mortality, both ICU and in-hospital, was similar in both groups (Table 2).
Table 2. Clinical parameters

<table>
<thead>
<tr>
<th></th>
<th>Rh-APC (n=16)</th>
<th>Placebo (n=11)</th>
<th>P-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>SOFA</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Day 1</td>
<td>8 (3)</td>
<td>9 (2)</td>
<td>.67</td>
</tr>
<tr>
<td>Day 5</td>
<td>8 (2.75)</td>
<td>8 (2)</td>
<td>.70</td>
</tr>
<tr>
<td>Day 15</td>
<td>4.5 (4.75)</td>
<td>8 (5)</td>
<td>.12</td>
</tr>
<tr>
<td>SAPS</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Day 1</td>
<td>47 (28.5)</td>
<td>36 (25)</td>
<td>.32</td>
</tr>
<tr>
<td>Day 5</td>
<td>38.5 (14.25)</td>
<td>34 (18)</td>
<td>.43</td>
</tr>
<tr>
<td>Day 15</td>
<td>31.5 (8)</td>
<td>35 (5)</td>
<td>.24</td>
</tr>
<tr>
<td>LIS</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Day 1</td>
<td>2.75 (1.5)</td>
<td>2.5 (0.5)</td>
<td>.12</td>
</tr>
<tr>
<td>Day 5</td>
<td>2.1 (1.2)</td>
<td>2.75 (1.7)</td>
<td>.58</td>
</tr>
<tr>
<td>Day 15</td>
<td>1.3 (1.4)</td>
<td>2.75 (3.25)</td>
<td>.92</td>
</tr>
<tr>
<td>Days on ventilator</td>
<td>8.5 (14)</td>
<td>8 (24)</td>
<td>.86</td>
</tr>
<tr>
<td>Ventilator-free days</td>
<td>11 (22)</td>
<td>11 (24)</td>
<td>.75</td>
</tr>
<tr>
<td>28-day mortality</td>
<td>3</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>ICU mortality</td>
<td>4</td>
<td>5</td>
<td>.41</td>
</tr>
<tr>
<td>In-hospital mortality</td>
<td>5</td>
<td>5</td>
<td>.69</td>
</tr>
<tr>
<td>SOFA</td>
<td>8 (3)</td>
<td>9 (2)</td>
<td>.67</td>
</tr>
</tbody>
</table>

Data are expressed as medians (interquartile ranges), or absolute numbers where appropriate.

Rh-APC, Recombinant human activated protein C; SOFA, Sequential Organ Failure Assessment; LIS, Lung Injury Score; SAPS, Simplified Acute Physiology Score; ICU, intensive care unit.

**DISCUSSION**

We have analyzed the effect of systemically administered rh-APC on pulmonary coagulopathy in patients with ARDS. The results can be summarized as follows: (a) infusion of rh–APC increased APC levels in the pulmonary compartment in patients with ARDS, (b) infusion of rh–APC attenuated systemic coagulopathy, (c) and pulmonary coagulopathy in patients with ARDS, (d) infusion of rh–APC resulted in faster resolution of pulmonary dysfunction, and (e) infusion of rh–APC was not associated with bleeding complications.

Reduced levels of (activated) protein C are associated with non-pulmonary organ system dysfunction and increased mortality in ARDS [11]. Equally, high levels of PAI-1 prognosticate a poor outcome [11,24]. Although it has been suggested that APC may have beneficial effects in ARDS [5,25], as demonstrated in numerous animal models of ARDS, and in a human model with intrapulmonary delivered endotoxin (LPS) [9], this is the first study to investigate this hypothesis in clinical patients. Even though rh-APC is no longer licensed for the treatment of severe sepsis, it still may have a role in the treatment of ARDS.

APC is a natural anticoagulant that inactivates coagulation factors Va and Vllla. Furthermore, it decreases the synthesis and expression of sTF on leukocytes [26,27]. Systemic administration of rh-APC expectedly resulted in increased plasma levels of APC during the infusion period of 96
hours after which plasma levels of APC returned to levels comparable to the placebo group. In line with this finding, as has been demonstrated previously, systemic levels of TATc and PAI-1 decreased while PAA increased [28].

Levels of TATc and sTF have previously been used as markers for local coagulation activation [9]. At baseline, concentrations of TATc and sTF were comparable in the rh-APC and placebo group, indicating a comparable amount of coagulation activation. The plasma levels of both TATc and sTF in NBLF were markedly reduced after the infusion of rh-APC. In addition, APC was detectable in NBLF of these patients and still increased as compared to baseline on day 5, when rh-APC infusion had already been stopped. This is an interesting finding, since APC was no longer detectable in the blood within 2 hours after the infusion was stopped in the majority of patients with sepsis [29]. This suggests that APC can enter the alveolar space after intravenous administration and that its pulmonary clearance is relatively slow.

During sepsis, systemic fibrinolysis is inhibited mainly as a result of elevated levels of PAI-1[30]. In the alveolar compartment of patients with ARDS, fibrinolysis is suppressed as well [31]. APC may augment fibrinolysis through inhibition of PAI-1. Our results show that APC markedly reduced PAI-1 levels in NBLF. PAA however, was enhanced systemically compared to baseline, but this enhancement did not reach statistical significance in NBLF. A possible explanation may be that tissue-type plasminogen activator (tPA) and urokinase-plasminogen activator (uPA), together responsible for the largest part of PAA, are bound to their inhibitor PAI-1. That way, the net PAA may have been decreased because of heightened formation of PAI-1. Comparable observations were made in the alveolar compartment of patients with pneumonia, and in healthy volunteers challenged with intravenous or intrapulmonary LPS [9,32,33].

The present study on the effects of rh-APC on pulmonary coagulopathy in patients with ARDS, is in line with previous findings in sepsis and a human model of lung injury [9,34]. Systemic administration of rh-APC results in reduced activation of coagulation both systemically and in the alveolar compartment. These anticoagulant effects are exerted through inactivation of factors Va and Vlla, and as a consequence inhibition of thrombin formation. This may in part be enhanced by inhibition of TF, which may have prevented protein C consumption [27]. A profibrinolytic effect of APC could not be demonstrated in the alveolar compartment, as opposed to the systemic effects. Our findings are in conflict with two earlier studies investigating the effect of rh-APC, administered in a similar fashion as in our study, in human volunteers intravenously challenged with LPS, which did not observe an effect of the treatment on coagulation activation [35,36]. In these studies, however, only systemic markers of coagulation were measured. Furthermore, systemic coagulopathy caused by endotoxin in healthy volunteers may differ from systemic coagulopathy in critically ill patients.

The aforementioned effects, may well have resulted in the faster resolution of pulmonary dysfunction in the patients treated with rh-APC, as expressed by the more rapid decrease in LIS, as compared to patients treated with placebo. The LIS consists of PaO2/FIO2-ratio, level of PEEP,
compliance, and pulmonary radiography. These single components improved over time, although the difference between both treatment groups did not become statistically significant. Yet, this did not translate into a reduced number of ventilator-free days, compared to the placebo group, which may be attributable to the relatively small sample size.

Our study has some important limitations. We chose to analyze the effects of this trial on systemic and pulmonary coagulation only in intubated and mechanically ventilated patients in which it was deemed safe to perform serial NBL, thereby inducing selection bias. We may have missed out on the more severe cases of ARDS, which factor on the other hand may be balanced by the patients that were not sedated enough to undergo NBL. Also, the relatively small study populations may have resulted in a limited statistical power to detect clinical differences between study groups.

Furthermore, the study population of our trial differed significantly from that of most clinical trials of ARDS because of exclusion of those patients with severe sepsis or septic shock who had an indication for rh-APC treatment in line with international guidelines at the time of the study, and with the exclusion of patients with increased risk of bleeding. Indeed, we had to exclude 94% of patients with ALI. This is, however, in line with a clinical trial comparing rh–APC with placebo with similar in– and exclusion criteria [37].

Finally, this trial was initiated when rh-APC was still in use. Nowadays, commercial rh-APC (Xigris®) has been taken off the market, and its role in the treatment of severe sepsis appears to have subsided, although controversies remain [38]. Our trial however, may contribute identifying new areas in which rh-APC can possibly be of value in the near future.

**Conclusion**

This study demonstrates for the first time in selected mechanically ventilated patients with ARDS that systemically administered rh-APC has a powerful pulmonary anticoagulant effect. In patients treated with rh-APC, this resulted in a faster resolution of pulmonary dysfunction as expressed by an improvement of LIS, and organ dysfunction in general as expressed by an improved SAPS. Together with previous (animal) studies, the present study underlines the potential of APC as a therapy for ARDS.

**Disclosure**

This study was partly funded by Eli Lilly, Indianapolis, IN, USA. Eli Lilly provided the study medication and financed the execution of this study.
REFERENCES


Activated protein C attenuates pulmonary coagulopathy in patients with acute respiratory distress syndrome | 121


